

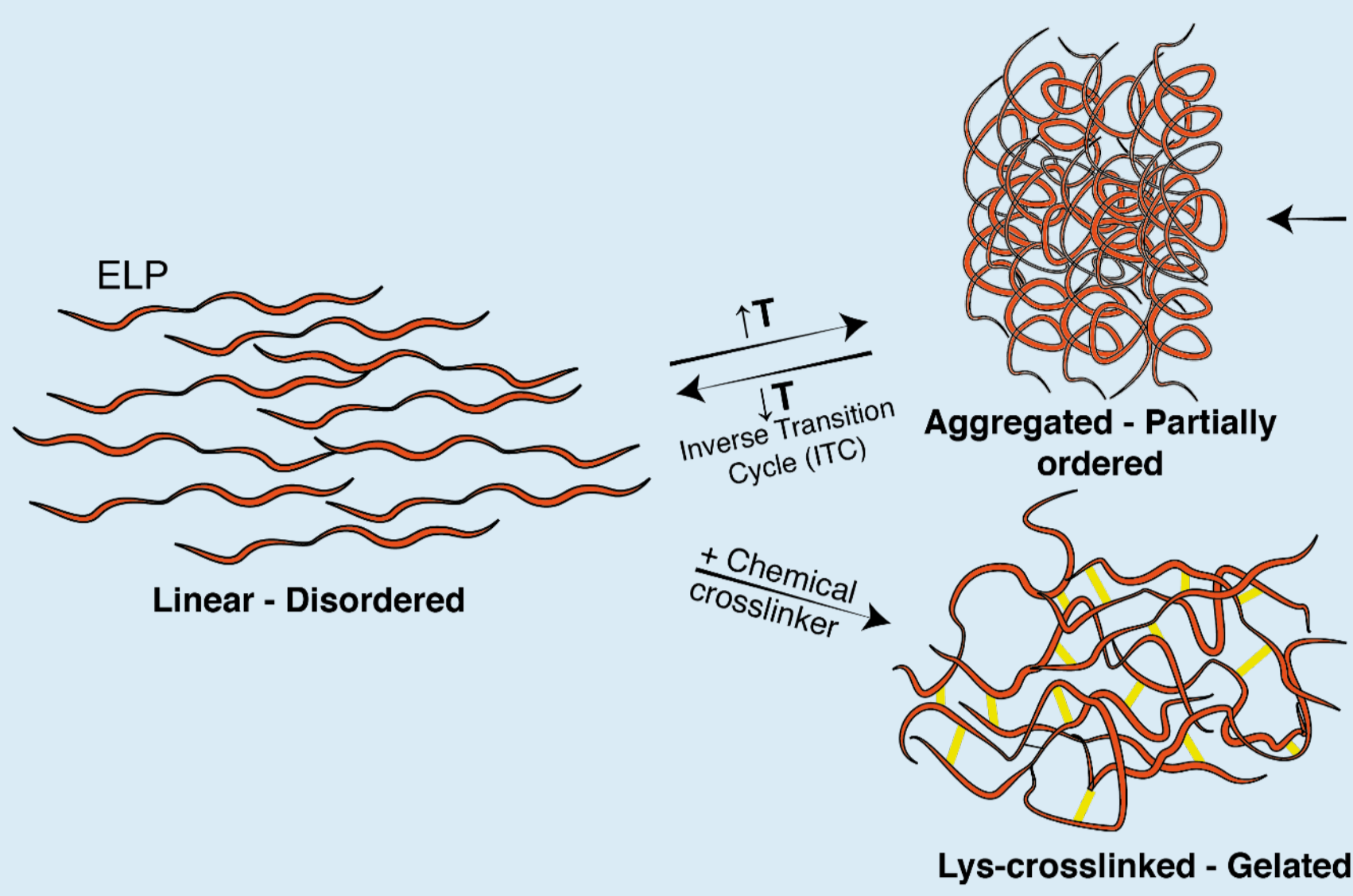


ON THE SYNTHESIS AND STRUCTURAL CHARACTERISATION OF ELASTIN-LIKE RECOMBINAMER HYDROGEL SCAFFOLDS FOR TISSUE REGENERATION

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INTRODUCTION



Elastin-like polypeptides (ELPs) are applied in a broad range of biomedical fields, such as drug delivery and tissue engineering. These genetically engineered peptides are designed to mimic the endogenous elastin and are characterised by a repeating (VPGXG)_n motif, where X could be any guest residue except for proline. Due to the periodic occurrence

of *cis*-proline residues, such peptides exhibit a partially ordered behaviour, undergoing reversible transitions from a disordered state to a β -spiral secondary structure. This is known as Inverse Transition Cycle (ITC) and occurs under a specific temperature, related to the peptide primary structure. Incorporation of lysine residues in low frequency, provides the ability of chemical crosslinking, thus forming a nanostructured hydrogel of good mechanical properties, able to function as biomaterial for tissue scaffolding.

CHARACTERISATION

SEM

- Surface visualisation
- Topography

TEM

- Bulk characterisation
- Pore size

AFM

- Surface topography
- Mechanical properties (Stiffness, Young's Modulus)

Molecular Dynamics Simulations

- Coarse Grained model
- Aggregation mechanism
- Structural transitions - Inverse Transition Cycle
- *In silico* evaluation step of experimental results

CURRENT RESULTS

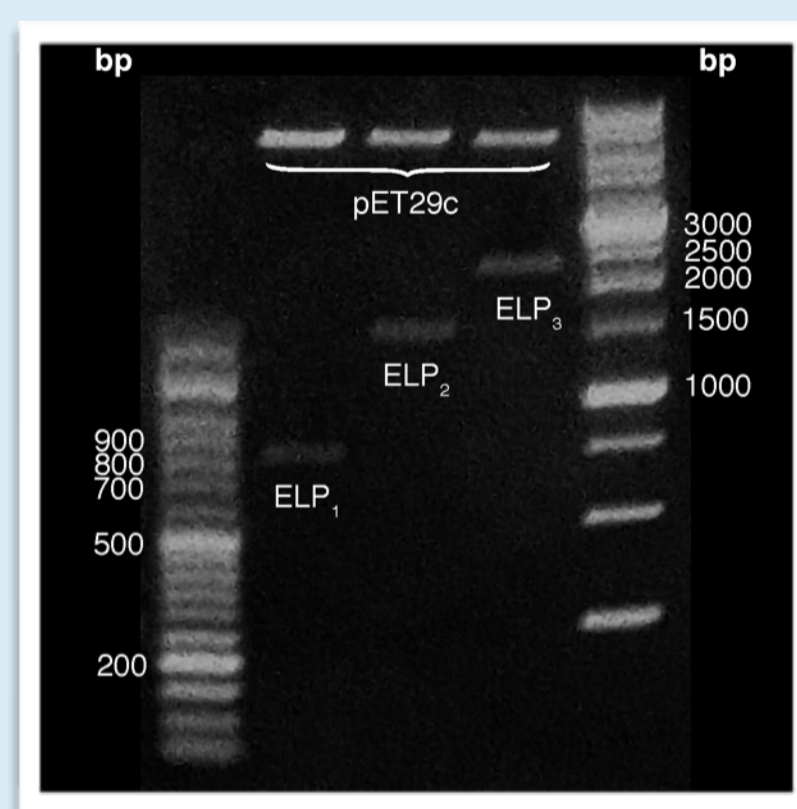


Figure 3: NdeI/KpnI digestion of plasmids containing one, two and three NAV-ELP gene repeats

• We have successfully constructed two ELP genes (one containing the NAV motif and one control peptide without it) by the methods described (Figure 3).

• Sequencing results show consistency with the predicted DNA sequence.

• The NAV-ELP and control-ELP constructs, code for peptides of ~57kDa and ~49kDa respectively.

• We are currently making attempts to optimise overexpression conditions and achieve maximum yield of the ELP peptides (non-avidin and control peptide).

• Overexpression conditions so far (see western blot at Figure 4):

- ➔ NAV-ELP: LB Broth-kanamycin, 37°C, 120rpm, 4h, +1mM IPTG at OD₆₀₀=0.4
- ➔ CTRL-ELP: LB Broth-kanamycin, 37°C, 120rpm, 4h, +1mM IPTG at OD₆₀₀=0.6

• Further optimisation is needed to eliminate proteolysis (multiple bands in the western blot)

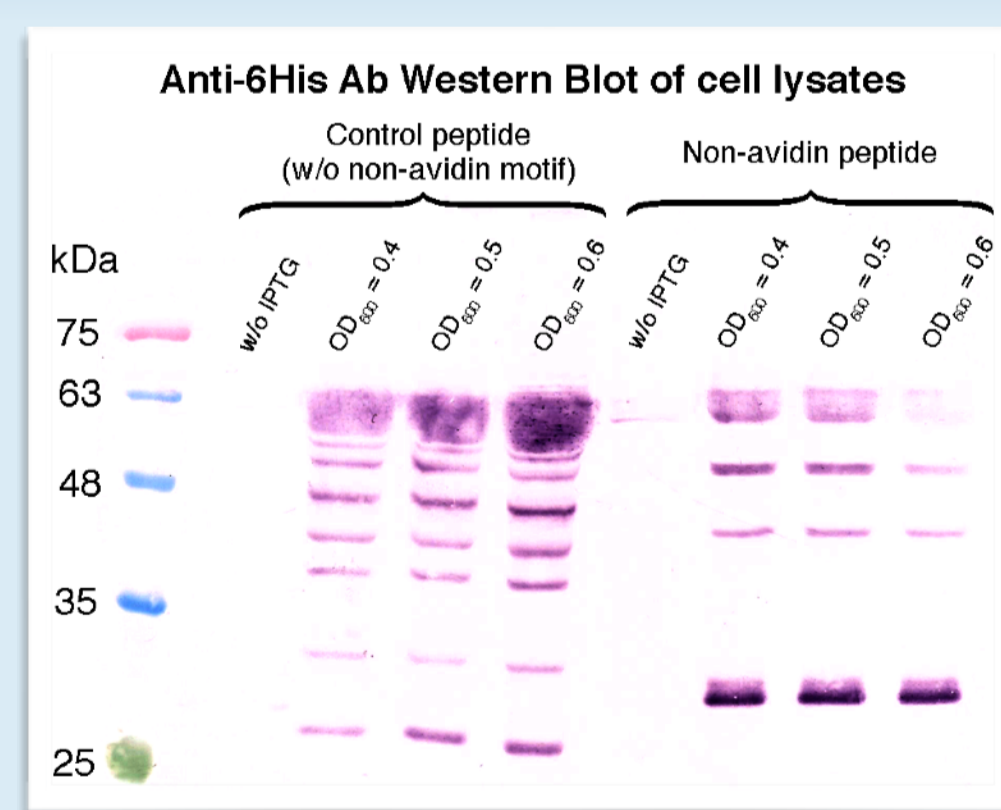


Figure 4: ELP peptide Western blot for semi-quantitative assessment of expression efficiency at different induction times

METHOD OUTLINE

ELP genes design and synthesis (vector: pET29c, host: TOP10 E.coli)

Overexpression in BL21 E.coli (after optimization of conditions)

ELP purification (ITC protocol and/or Ni-NTA column)

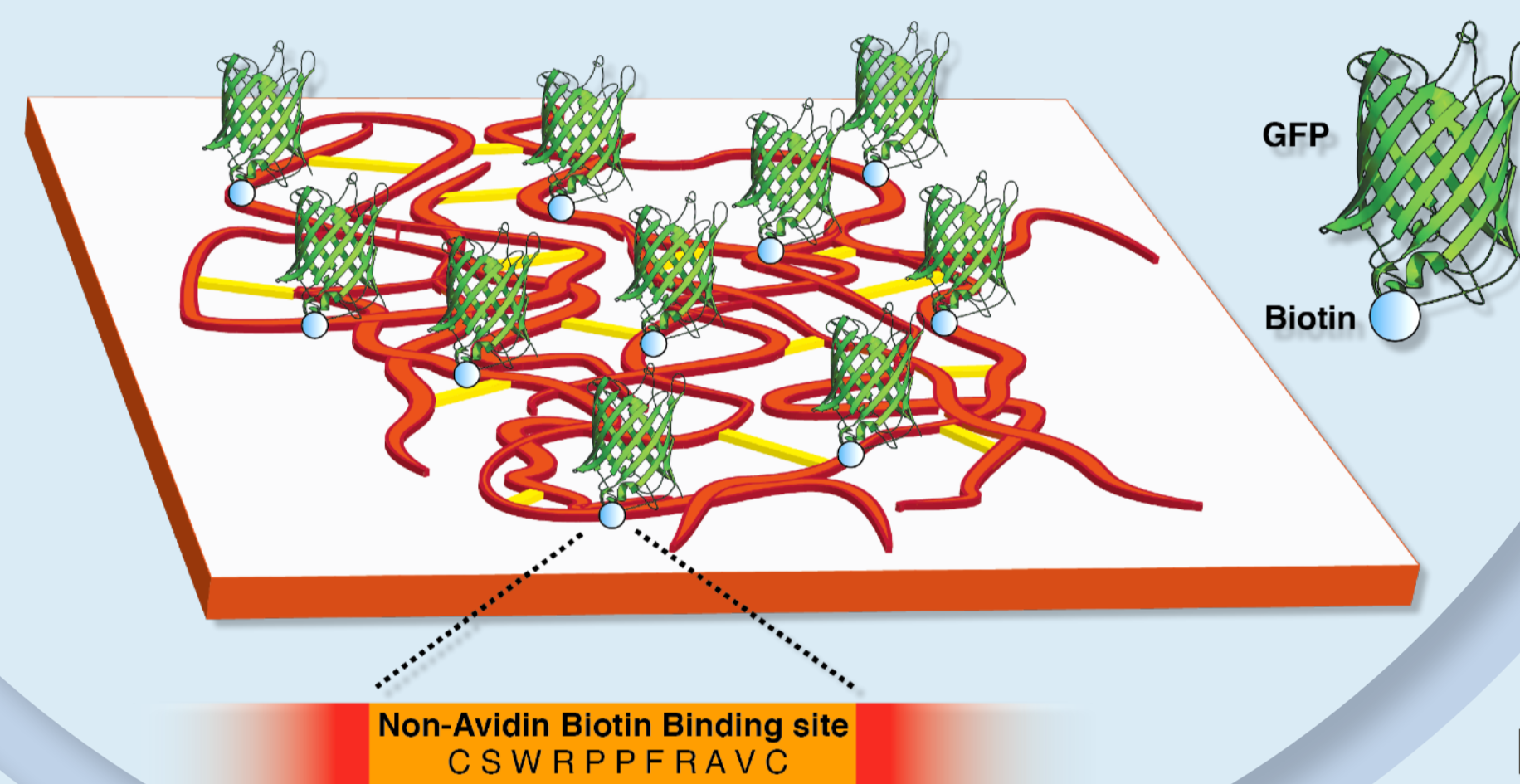
Chemical crosslinking - hydrogel formation

Test with biotinylated GFP

Growth factor docking - cell seeding

AIM

- Fabrication of a customisable, biocompatible hydrogel scaffold for cartilage and bone regeneration
- Genetically synthesised ELPs as biopolymeric material
- Controlled incorporation of biotinylated proteins or peptides on the ELP molecules (e.g. biotinylated GFP for testing, see figure below) through a Non-Avidin-related (NAV) biotin binding motif.
- Docking of biotinylated growth factors to promote differentiation and proliferation of progenitor cells



GENE SYNTHESIS

• Artificial ELP genes were *ab initio* synthesised through a PCR-based strategy (also known as Recursive PCR, Prodromou et al), which involves multiple partially-overlapping primers (one forward and several reverse ones) and no template DNA. Figure 1 schematically summarises the method. This a single-step procedure which does not require multiple PCR reactions to produce the final product.

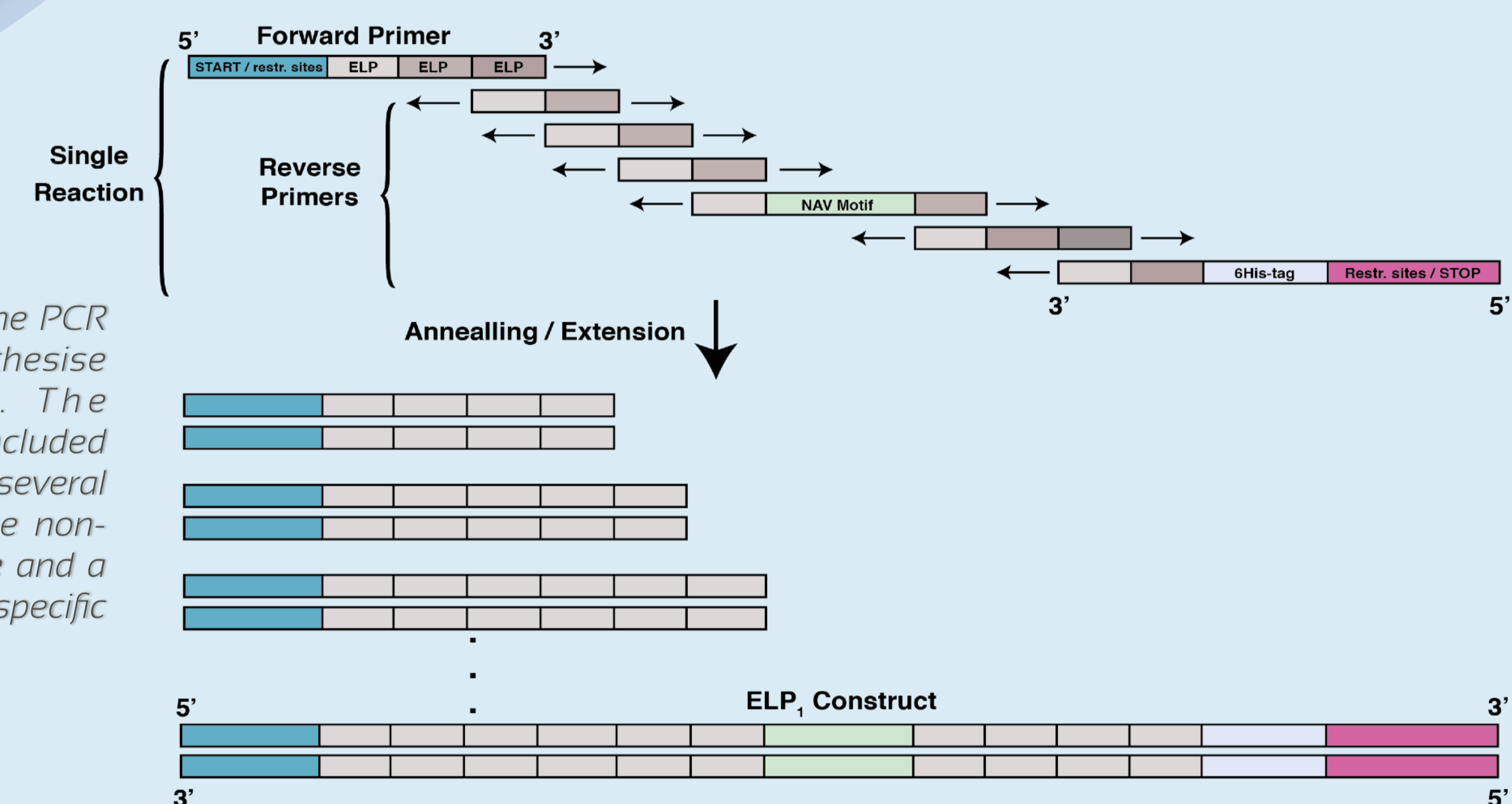


Figure 1: Schematic of the PCR method used to synthesise initial ELP genes. The component primers included start and stop codons, several ELP tandem repeats, the non-avidin biotin binding site and a 6His-tag as well as specific restriction sites.

• Two additional repeats of the whole gene synthesised by the above method were introduced, using a method known as Recursive Directional Ligation (RDL) (Figure 2). Two ELP₁ monomers were cut from their vector plasmids, one on the 5' and one on the 3' end, and ligated together to form an ELP₂ dimer. One more repeat was introduced in a similar way to form the final ELP₃ construct. TOP10 E. coli strains were used as hosts.

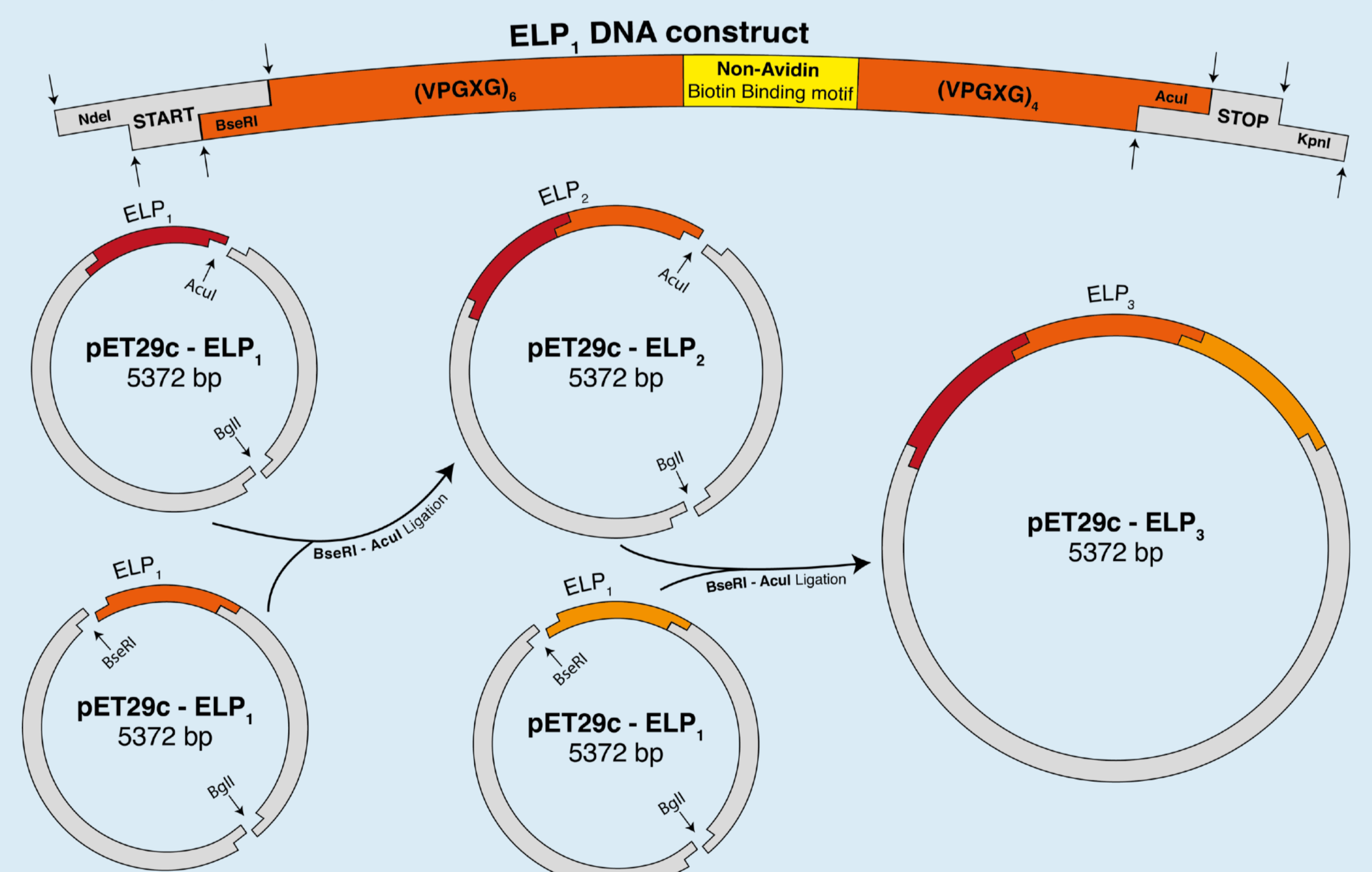


Figure 2: Schematic of the Recursive Directional Ligation method, used to insert two extra tandem repeats of the initial ELP gene. Note the restriction enzyme combinations for each step of the procedure.

REFERENCES

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